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enclose herewith a declaration of Dr. Jeffrey D. Milbrandt and an Information Disclosure Statement with copies of the references that are cited in the declaration and in this paper.

### REMARKS

Claims 1 and 5-9 are pending. Claims 5 and 6 were cancelled without prejudice in the previous response, filed on June 10, 2002. Therefore, claims 1 and 7-9 are presently pending in the instant application.

Claims 1 and 7-9 stand rejected under 35 U.S.C. § 112, first paragraph, for an alleged failure to provide a description sufficient to enable a person of ordinary skill in the art to practice the claimed invention. At p. 3-4 of the office action, the Examiner contends that the instant specification fails to provide any evidence or sound scientific reasoning that would support a conclusion that the claimed chimeric peptides of SEQ ID NO:23 and 26 can activate GFR $\alpha$ 1-RET, but does not substantially activate GFR $\alpha$ 2-RET or GFR $\alpha$ 3-RET. The Examiner reasoned that while the examples in the specification use murine chimera peptides, the biological activity of the claimed human chimeric peptides "was never tested".

Applicants respectfully assert that the specification of the instant application enables a person of ordinary skill in the art to practice the full scope of the claims. Specifically, the specification of the instant application teaches a person of ordinary skill in the art that the human PSPN chimera, like the mouse PSPN chimera, can activate the GFR $\alpha$ 1-RET but does not substantially activate GFR $\alpha$ 2-RET or GFR $\alpha$ 3-RET. A person of ordinary skill in the art would reasonably conclude that the human PGP-F2ac (SEQ ID NOs: 23 and 26) can be readily substituted for the mouse PGP-F2ac in the same assays described in the specification (Examples 3, 4, and 5), and the human PGP-F2ac would yield the same result or substantially the same result. Therefore, repetition of the examples in the instant application with human PGP-F2ac lacks scientific justification and would be redundant. In support of Applicants' arguments, Applicants submit herewith a declaration of Dr. Jeffrey D. Milbrandt.

First, the specification of the instant application clearly demonstrates that a PGP-F2ac chimera can activate GFR $\alpha$ 1-RET but does not substantially activate GFR $\alpha$ 2-RET or GFR $\alpha$ 3-RET. As shown in Figure 6 of the instant application, a wild-type PSPN cannot activate GFR $\alpha$ 1-RET. As demonstrated in Example 3, after substitution of the F2a and F2c regions of GDNF

into PSPN, the PSPN chimera can activate GFR $\alpha$ 1-RET. Example 4 shows that the PGP-F2ac cannot substantially activate GFR $\alpha$ 2-RET. As explained in the specification, artemin is the only GDNF family member that can activate GFR $\alpha$ 3-RET. (specification at p. 36, line 28). Example 4 shows that the Ha region in artemin confers the ability to activate the GFR $\alpha$ 3-RET. Without this region, as observed in PAP-F2ac (which behaves virtually identically to PGP-F2ac in activating GFR $\alpha$ 1-RET but not GFR $\alpha$ 2-RET; please see Figure 6), the chimera cannot substantially activate GFR $\alpha$ 3-RET. The claimed PGP-F2ac also does not contain an Ha region from artemin and therefore, a skilled artisan would reasonably expect that it cannot substantially activate GFR $\alpha$ 3-RET.

Second, while the Examiner is correct that in some instances, the primary structure of a protein cannot always predict its function, and a single amino acid change can alter the protein's function, in this case, the physical structure of the claimed human chimeric polypeptides reasonably correlates with its predicted function (i.e. they can activate the GFR $\alpha$ 1-RET, but do not substantially activate GFR $\alpha$ 2-RET or GFR $\alpha$ 3-RET). The chimeric polypeptides in the examples consist of mouse PSPN with substitutions in the respective F2a and F2c regions from GDNF. The amino acid sequence of the mouse PSPN is more than 85% identical to human PSPN and when conservative substitutions (*i.e.* those amino acids that share common chemical attributes and therefore have similar effects in determining the overall shape of the polypeptide) are considered, it is 94% similar to mouse PSPN. Further, the important F2a sequence (AFDDD) and F2c sequence (YHILRKH) from GDNF, which are incorporated into the chimera of the present application and which provide the GFR $\alpha$ 1 selectivity, are identical in GDNF derived from human, mouse, rat, zebrafish, and chicken. This is consistent with the observations that GFLs from different species have similar functions.

Consistent with the observation of a lack of species specificity in the functions of GFLs, decades of research show that other types of neurotrophic factors, which also exhibit species cross-reactivity, share conserved structural properties. For example, the neurotrophin family of neurotrophic factors (NGF, BDNF, etc.) demonstrates no species specificity in their function and share a highly conserved cysteine knot structure (as do GFLs). The original discovery of NGF resulted from experiments in which a mouse sarcoma was implanted into chicken embryos, whereby the NGF released from the mouse sarcoma produced a massive neural overgrowth of

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the neurons from the chicken host (*Levi-Montalcini*, R Ann. NY Acad. Sci, 55: 330-343 (1952)). The classical assay used to purify mammalian NGF was the chicken sensory neuron assay. Mouse salivary glands were identified as a preparative source of NGF after it was serendipitously observed that the NGF active on chicken sensory (and subsequently mammalian) neurons was present in snake venom *Cohen S.*, Proc Natl. Acad Sci USA 46:302-311 (1960). Like the neurotrophic factors, PSPN and other GFLs are rigidly held molecules due to its conserved cysteine knot structure generated by three internal disulfides within each monomer of the factors. Like the neurotrophic factors, these evolutionarily conserved molecules show virtually no species specificity in their actions.

Third, there has been no reported evidence of any species specificity among the GDNF family members. On the contrary, the evidence supports cross-reactivity among species. For example, the mouse PSPN-mouse NRTN chimera can activate human GFR $\alpha$ 2-RET and the mouse PSPN-human ARTN chimera can activate mouse GFR $\alpha$ 3-RET. In the present application, the PGP-F2ac was built by substituting into mouse PSPN the F2a and F2c regions from rat GDNF. As described above and in the specification, the mouse PSPN-rat GDNF chimera can activate rat GFR $\alpha$ 1-RET or human GFR $\alpha$ 1-RET without substantially activating GFR $\alpha$ 2-RET or GFR $\alpha$ 3-RET.

A wealth of literature supports the interchangeability of these factors, their respective GFR $\alpha$  coreceptors, and the Ret tyrosine kinase from a variety of species ranging from human to rodent to chicken. For example, in the initial description of the isolation and cloning of GDNF (*Lin et al.* Science 260: 1130-1132 (1993)), the authors showed that both rat and human GDNF promoted survival of rat dopaminergic neurons. In subsequent publications, human and rat GDNF was shown to promote neurite outgrowth and survival of chick sympathetic ganglion neurons (*Trupp et al.*, J Cell Bio 130:137-48 (1995)). Furthermore, human GDNF has been demonstrated to provide functional recovery in parkinsonian monkeys (*Gash et al.*, Science 380:252-255 (1996)). These examples are not restricted to GDNF, as NRTN also displays no species selectivity. Indeed, the original purification of NRTN from Chinese hamster ovary cells was monitored using a bioassay that utilized neurite outgrowth and survival of rat sympathetic neurons isolated from superior cervical ganglia (*Kotzbauer et al.*, Nature 384:467-470 (1996)).

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These data demonstrate the equivalence of members of this family from various species in a wide variety of assays as well as demonstrating the lack of any species-restricted functions.

Fourth, the original description of PSPN demonstrated that mouse PSPN was active in promoting survival of rat dopaminergic and motor neurons (*Milbrandt et al.*, Neuron 20:245-253 (1998)). It has been shown that PSPN-deficient mice suffer more severe damage to the cerebral cortex after ischemia (i.e. they have increased sensitivity to stroke) than wild type mice (*Tomac et al.* Proc Natl Acad Sci USA 99:9521-9526 (2002)). The authors found that treating these PSPN-deficient mice with human PSPN protected them from this increased susceptibility to ischemic damage. The neuronal damage caused by ischemia is the result of excessive glutamate release and subsequent elevation in intracellular calcium levels. The authors went on to show that mouse PSPN and human PSPN function virtually identically, with equivalent dose-response curves, to modulate glutamate-induced calcium influx in cortical neurons. Thus, mouse PSPN and human PSPN, despite their slight difference in primary sequence homology, have the same effects, both qualitatively and quantitatively, on protection from ischemia and modulation of intracellular  $[Ca^{2+}]$  levels.

In short, because members of this neurotrophic factor family, including human PSPN and mouse PSPN, do not show species specificity, and proteins from different species behave similarly in all assays examined thus far, the same GDNF sequences can be inserted into either the human or mouse PSPN to create the appropriate chimeras. The lack of species specificity and thus, the species interchangeability of the GFLs, are widely recognized in the field. For example, most investigators in the field using GDNF, NRTN, ARTN, or PSPN (or neurotrophins) do not even identify the species of origin when reporting the results of their experiments. Also, in *Tomac et al.*, Proc Natl Acad Sci USA 99:9521-9526 (2002), the authors performed almost all of their experiments using rhPSPN and did not bother to repeat the same experiments with rmPSPN even though their platform was a mouse model. Thus, a person of ordinary skill in the art would reasonably conclude that a chimera constructed using human PSPN would behave similarly to a mouse PSPN chimera in the same assay.

In light of the arguments presented above, the facts presented in the attached declaration, the teachings in the references cited in the declaration and submitted in the attached IDS, and the knowledge in the art, Applicants submit that the claims are fully enabled. Applicants


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respectfully request reconsideration and withdrawal of the rejection under 35 U.S.C. § 112, first paragraph.

### CONCLUSION

Applicants believe they have overcome or obviated all of the Examiner's rejections. Applicants submit that the pending claims are in proper form for allowance and respectfully request that such allowance be granted. If there are any outstanding issues that need to be resolved, the Examiner is invited to call the undersigned attorney.

Respectfully submitted,



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